



# Emerging technologies and tools for monitoring fish physiology and energetics



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## Today's contents

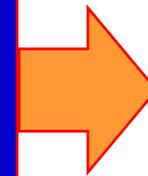
1. What should we monitor?
2. Artificial tags (biologging)
3. Natural tags (biogeochemical tags)
4. Combination with laboratory experiments and models
5. OceanDNA

# What should we monitor?

“Growth–mortality” hypothesis  
(Anderson 1988 JNAFS)  
survival rate is a function of growth  
rate and growth in the early life stage  
is the key for stock fluctuation

Fish migration has always played an  
essential role in marine conservation  
and fisheries management.  
(Moulton, 1939)  
Feeding & spawning migration

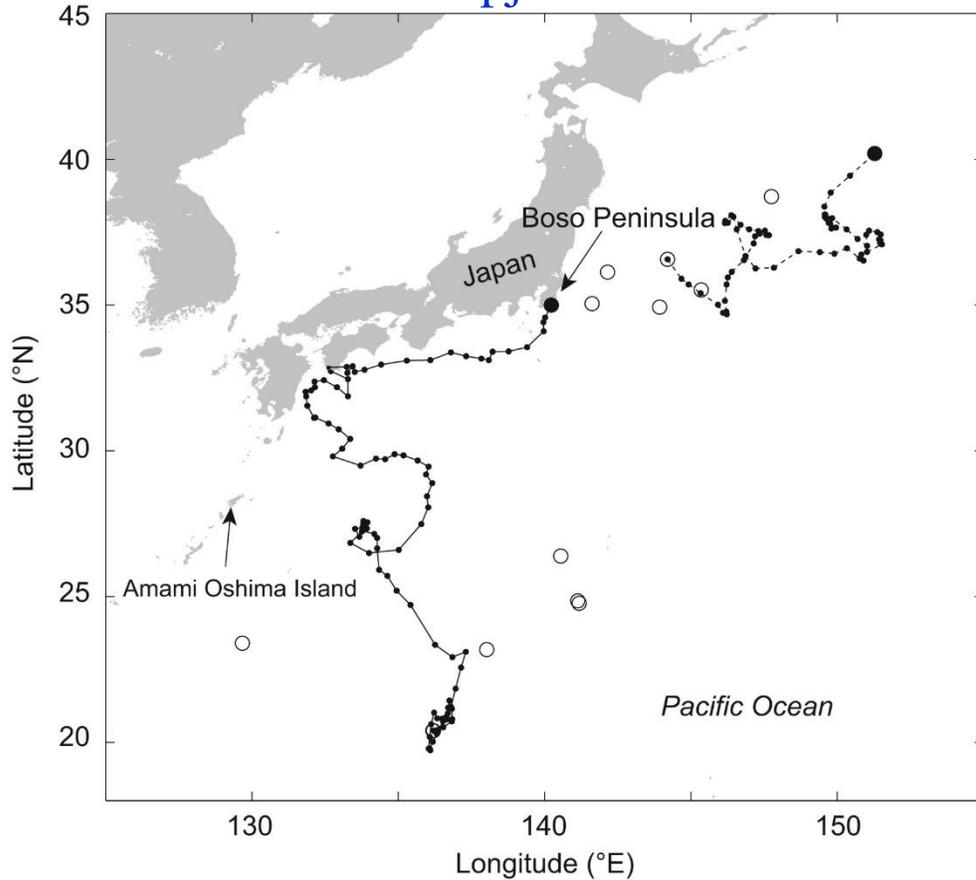
1. Environments fish experienced
2. Prey items fish predated
3. Growth of fish
4. Behaviors of fish
5. Migration route of fish
6. Distribution of fish



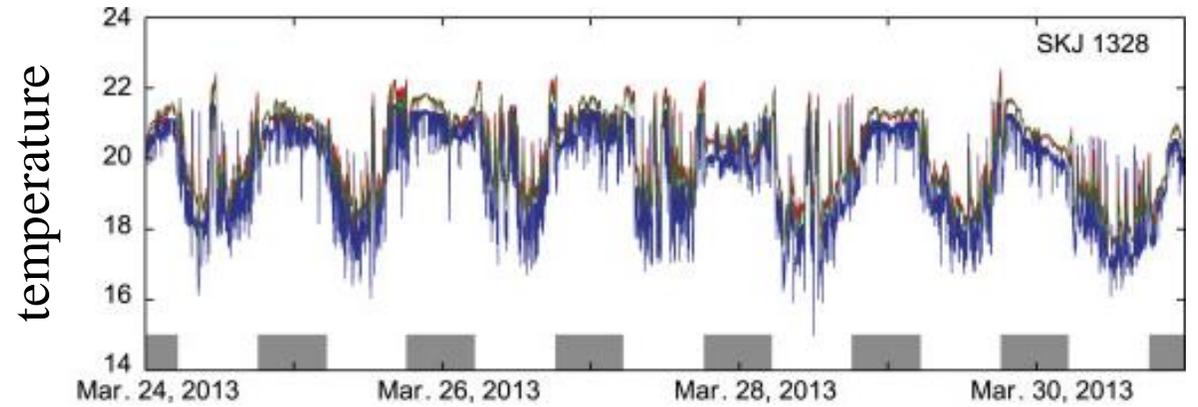
**Can investigate their  
life strategy**

# Artificial tags (biologging)

an example for archival tag observation  
of skipjack tuna



Aoki et al. (2017, DSR-II)



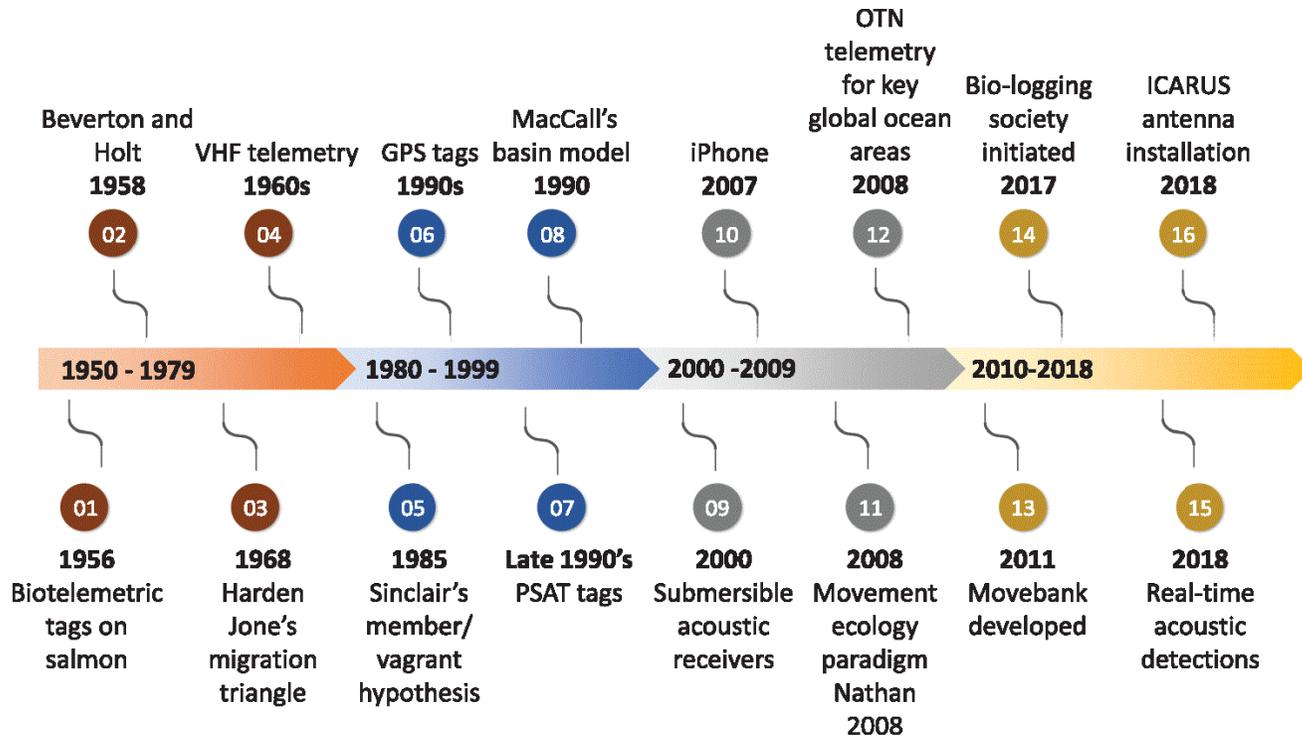
can record  
ambient temperature, body temperature,  
depth, light (position and moving speed),  
acceleration, video, etc.

an example for archival tag  
observation of salmon

Kitagawa et al. (2016)



# Artificial tags (biologging)



Lowerre-Barbieri et al. (2019)

- Biologging has been developed for long time and the applications have been extended.
- For larger fishes, biologging is the best approach to observe fish behaviors.

limitation  
 influence of tag for behaviors  
 position determination  
 lifetime of sensor  
 size

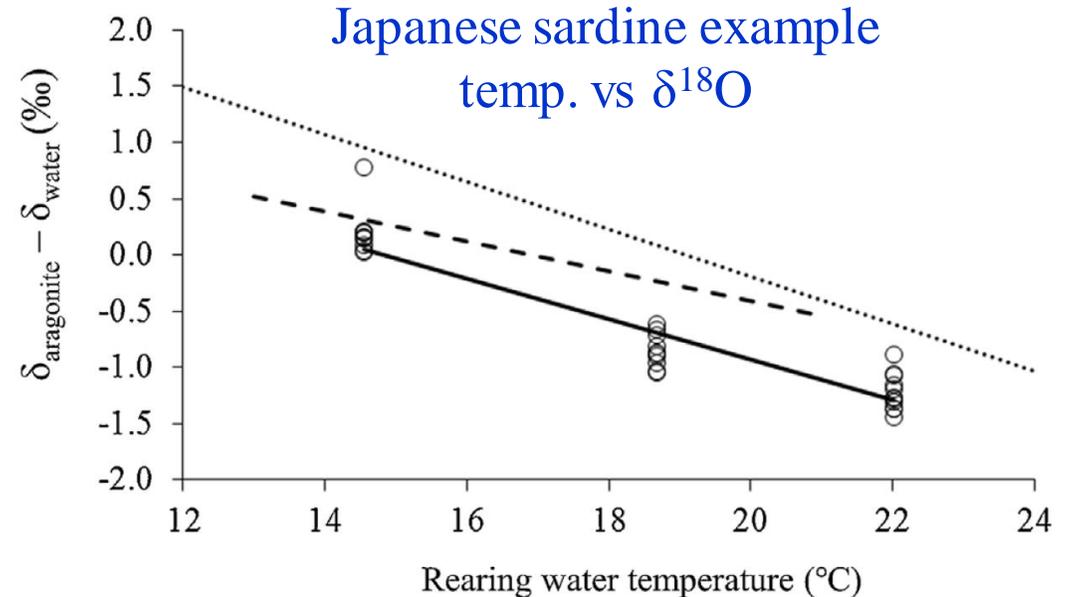
Application for smaller fish is still limited.  
 It is also impossible to apply to critical life stages (larvae & juvenile stages).

# Natural tags (otolith)

otolith of chub mackerel



Takahashi et al. (2013)



Sakamoto et al. (2017)

## Otolith

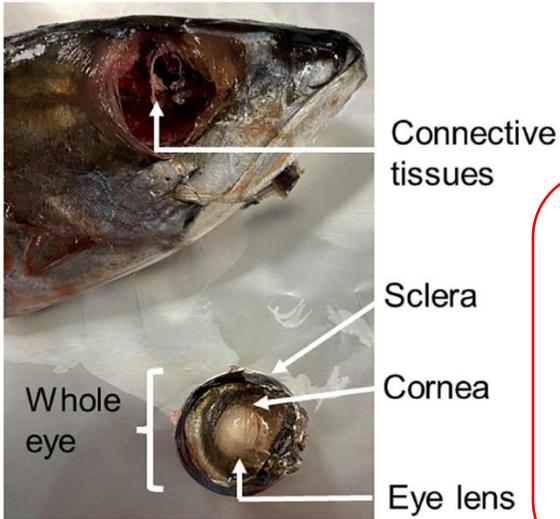
- a calcium carbonate structure in the inner ear.
- Daily rings are formed and growth of otolith indicates somatic growth.
- It is possible to estimate day age, if a hyaline zone is not formed.
- Chemical component analysis can be applied to reconstruct environmental history fish experienced.

# Natural tags (eye lens)

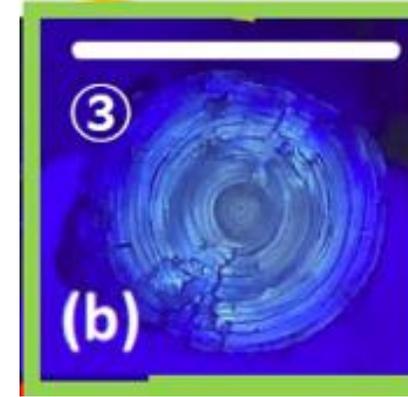
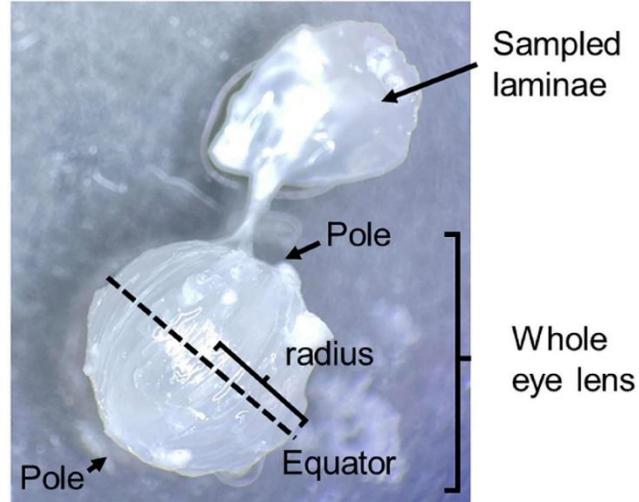
## 1. FISH MEASUREMENT



## 2. LENS EXTRACTION



## 3. DELAMINATION

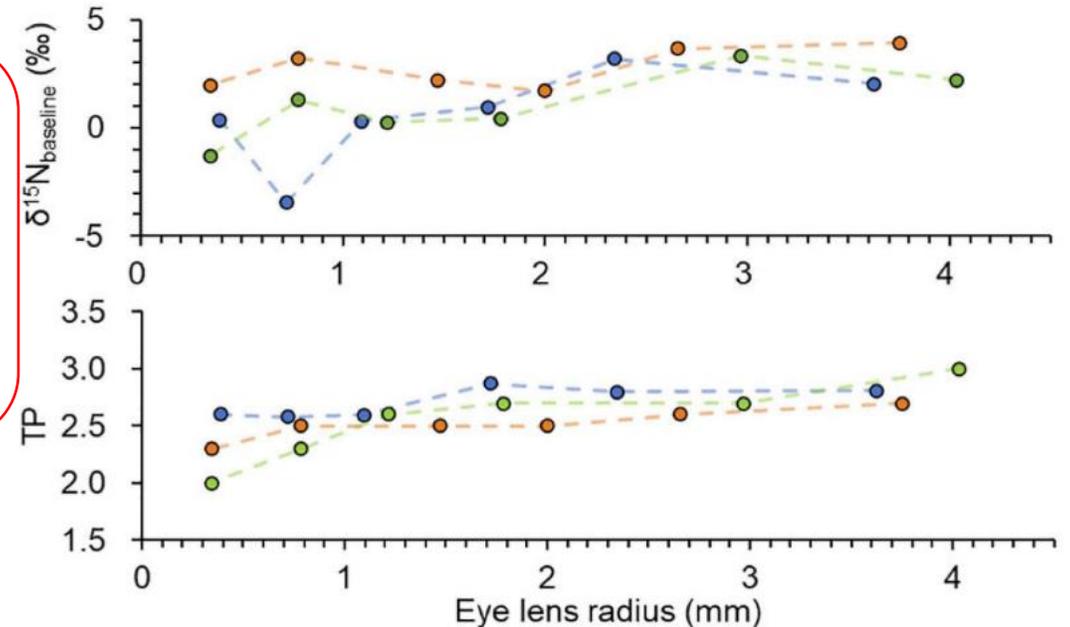


Quaeck Davies et al.  
(2018)

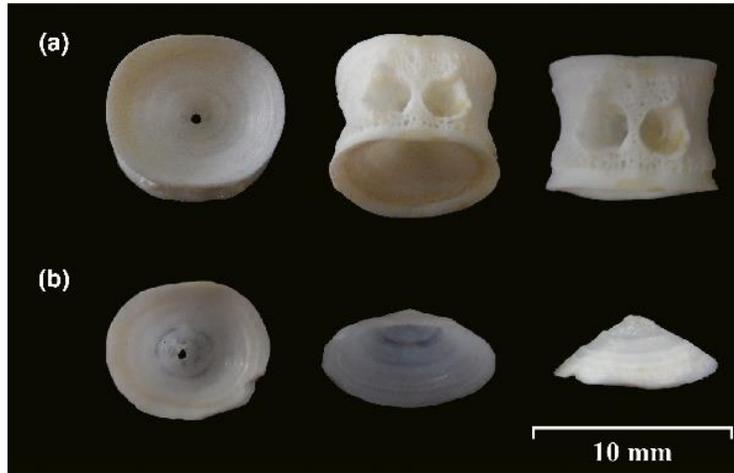
Eye lens  
( $\delta^{15}\text{N}$  of amino acids)

- migration history
- trophic level

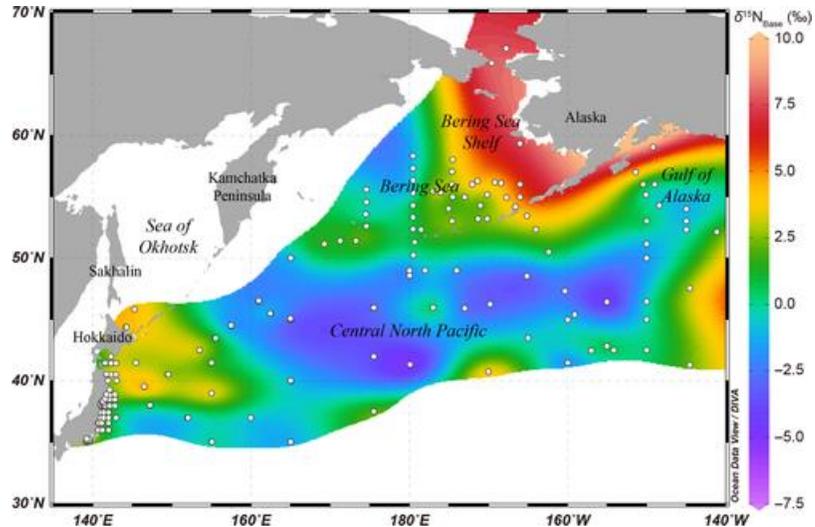
Harada, Ito, et al. (2022)



# Natural tags (bone: vertebral centra)



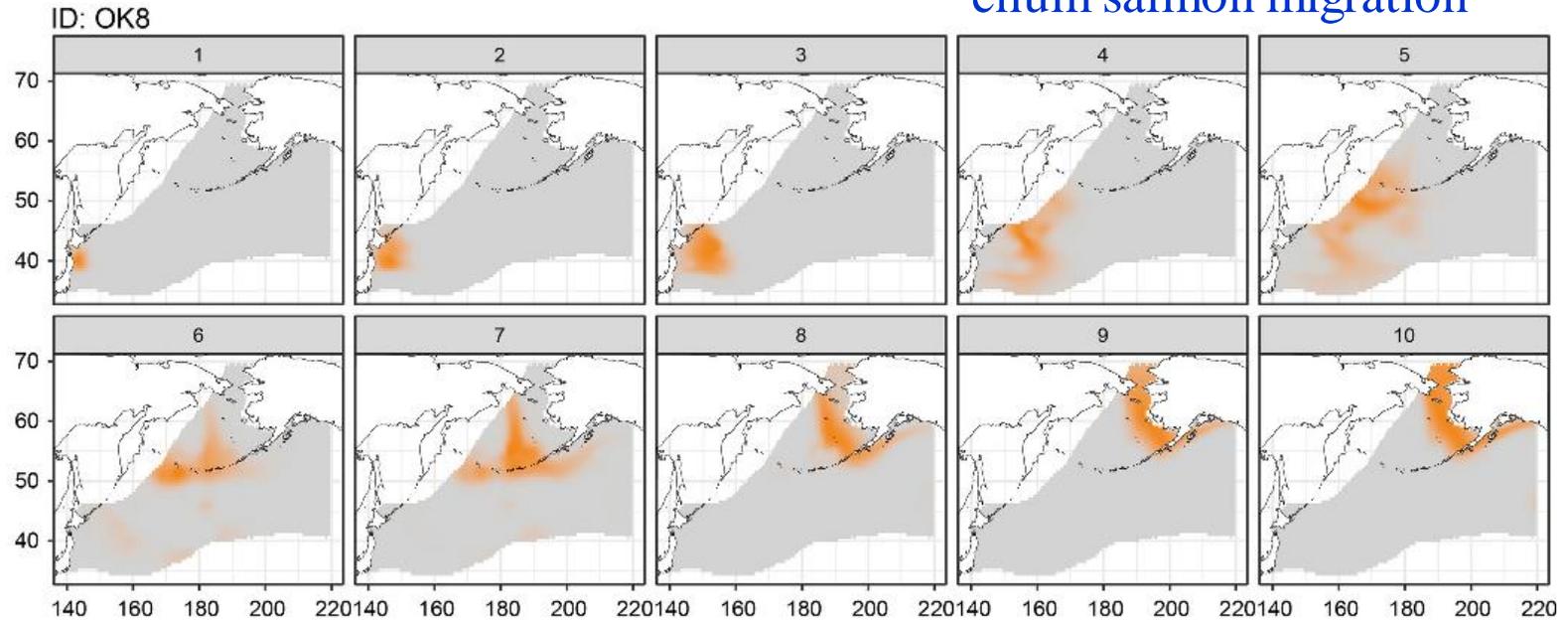
Matsubayashi et al. (2017)



isoscape of  
zooplankton  $\delta^{15}\text{N}$

Matsubayashi et al.  
(2020)

chum salmon migration

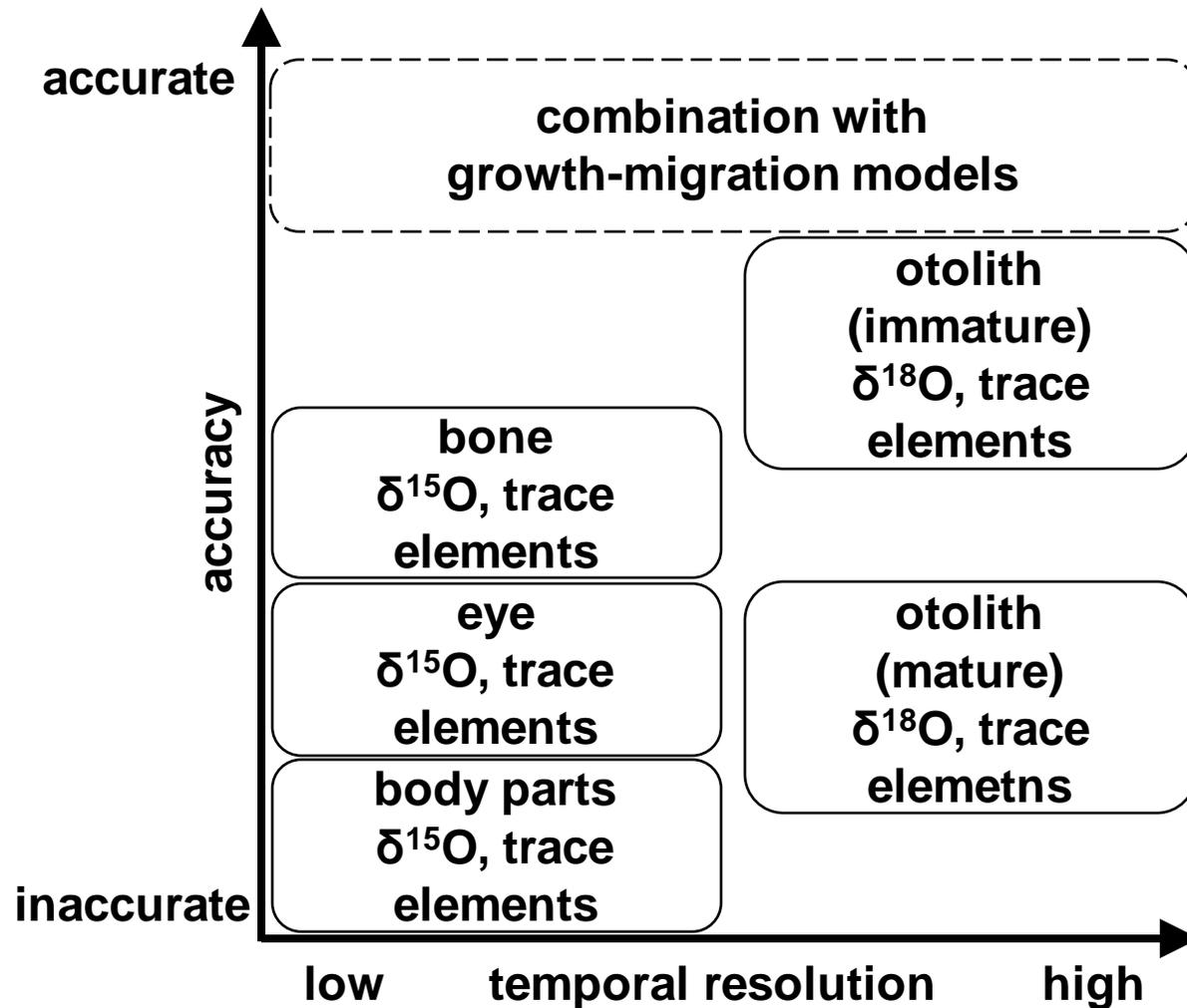


Matsubayashi et al.  
(2020)

Bone collagens  
( $\delta^{15}\text{N}$  of amino acids)

- migration history
- trophic level

# Natural tags



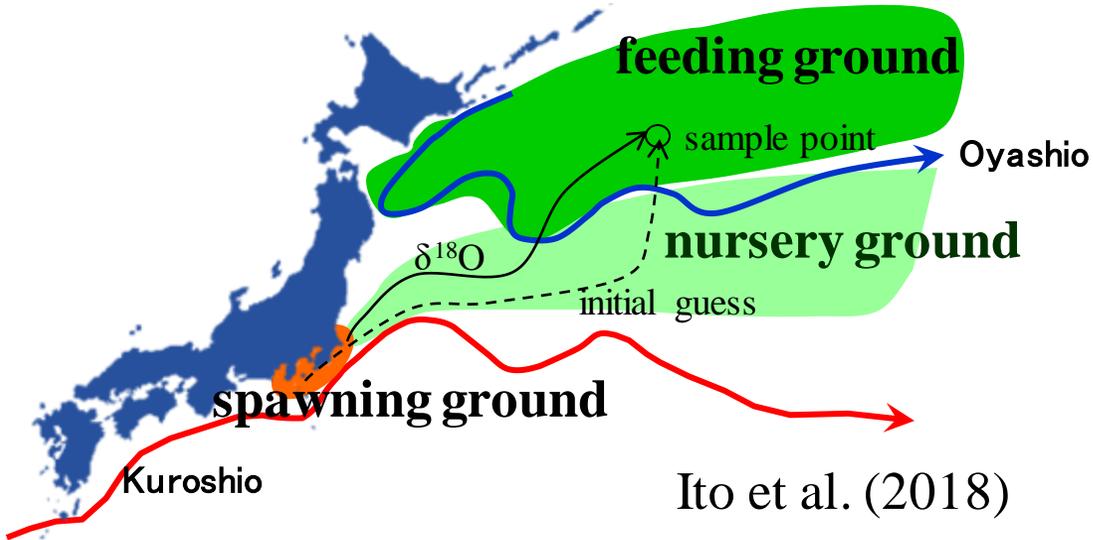
## strong points

1. no stress to fish
2. no size limitation
3. long record

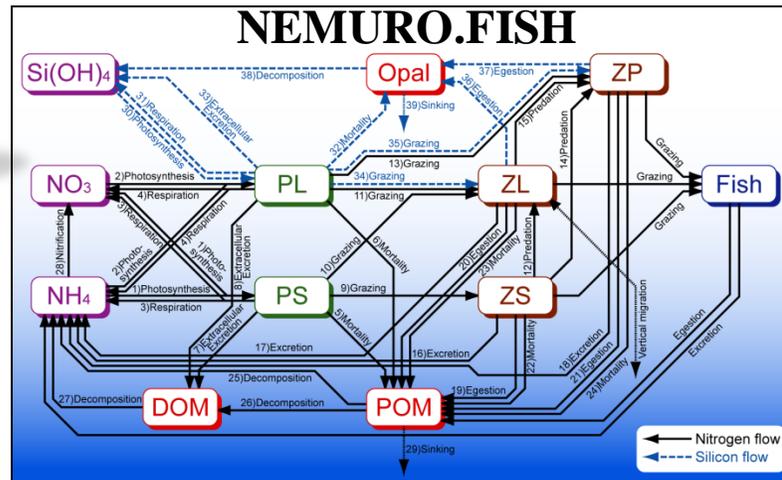
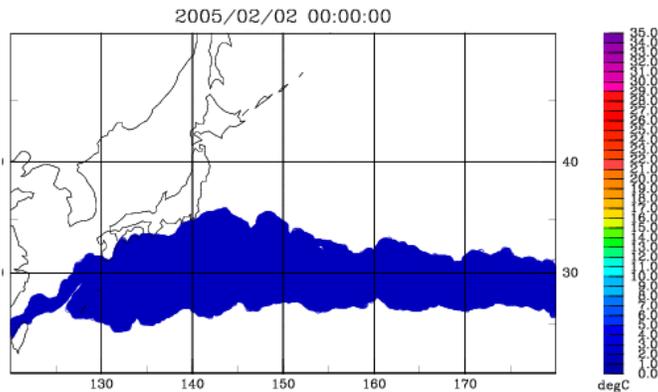
## weak points

1. limitation of temporal resolution
2. limitation of accuracy
3. limitation of spatial resolution

# Combination with fish growth-migration models



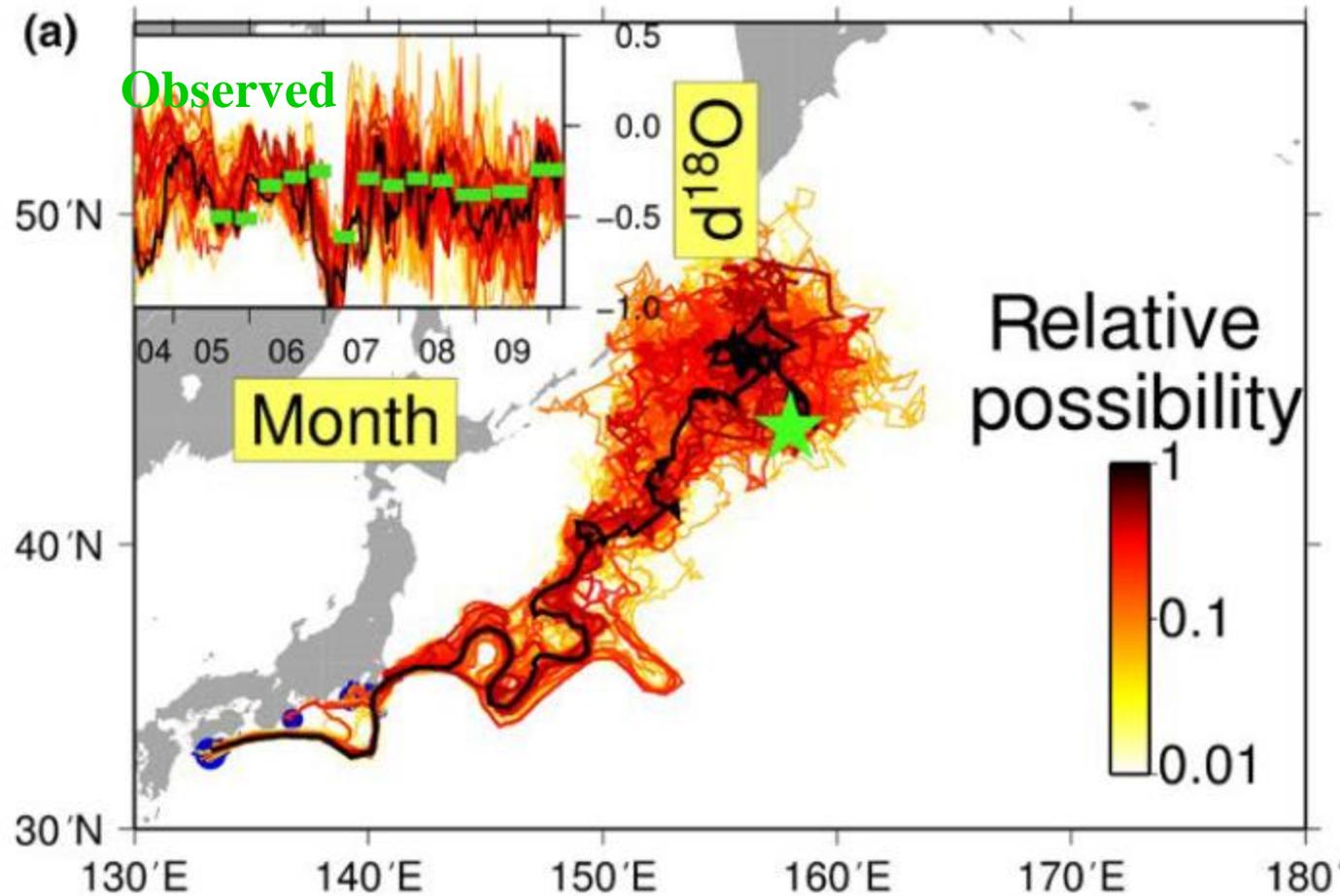
Combination of a fish growth-migration model (NEMURO.FISH) with natural tag chemical analysis enables precise estimation of migration routes and environments fish experienced. To build models, we need laboratory experiments.



Ito et al. (2004, 2007, 2010, 2013) etc.  
Megrey et al. (2007),

Guo, Ito et al. (2020, Fisheries Oceanography)  
Guo, Ito et al. (2021, Frontiers in Marine Science)

# Application for Japanese sardine



Sakamoto et al. (2018, Methods Ecol. & Evol.)

Early life migration history was estimated using otolith stable oxygen isotope history.

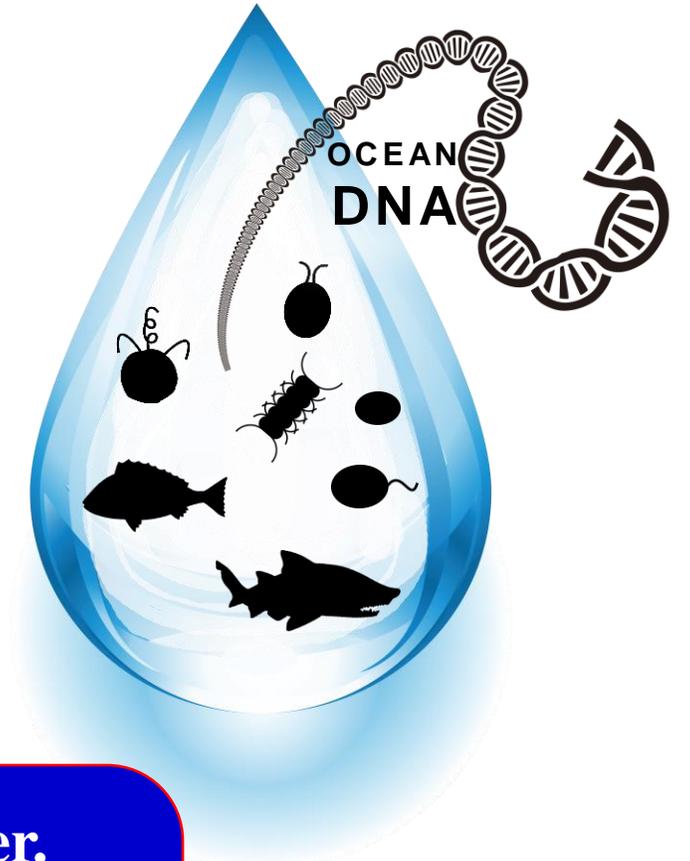
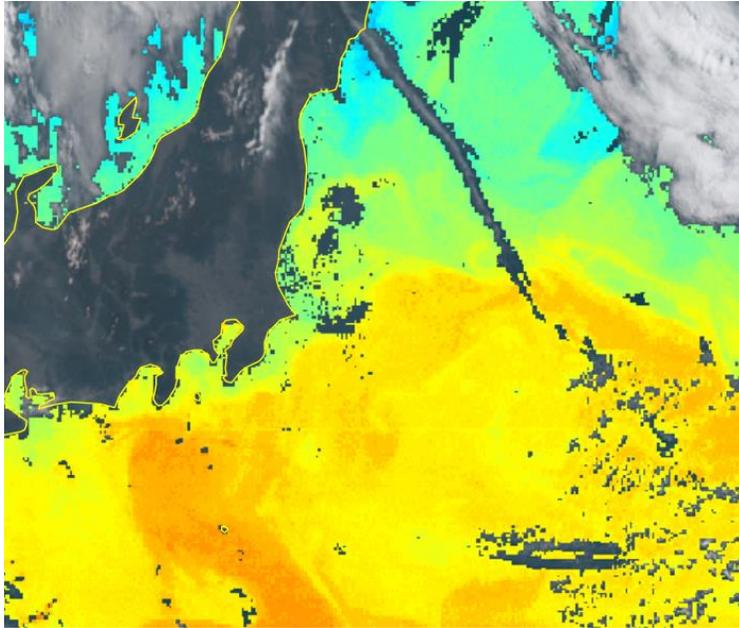
Temperature and salinity distribution estimated by a data assimilated ocean circulation model was used to estimate possible  $\delta^{18}\text{O}$  in otolith.

Random swimming fish was used to estimate possible migration route.

**Using model and natural tag chemical component analyses,  
fish migration route and experienced environment  
can be determined.**

**We want to confirm whether  
fish exist on the estimated migration route or not.**

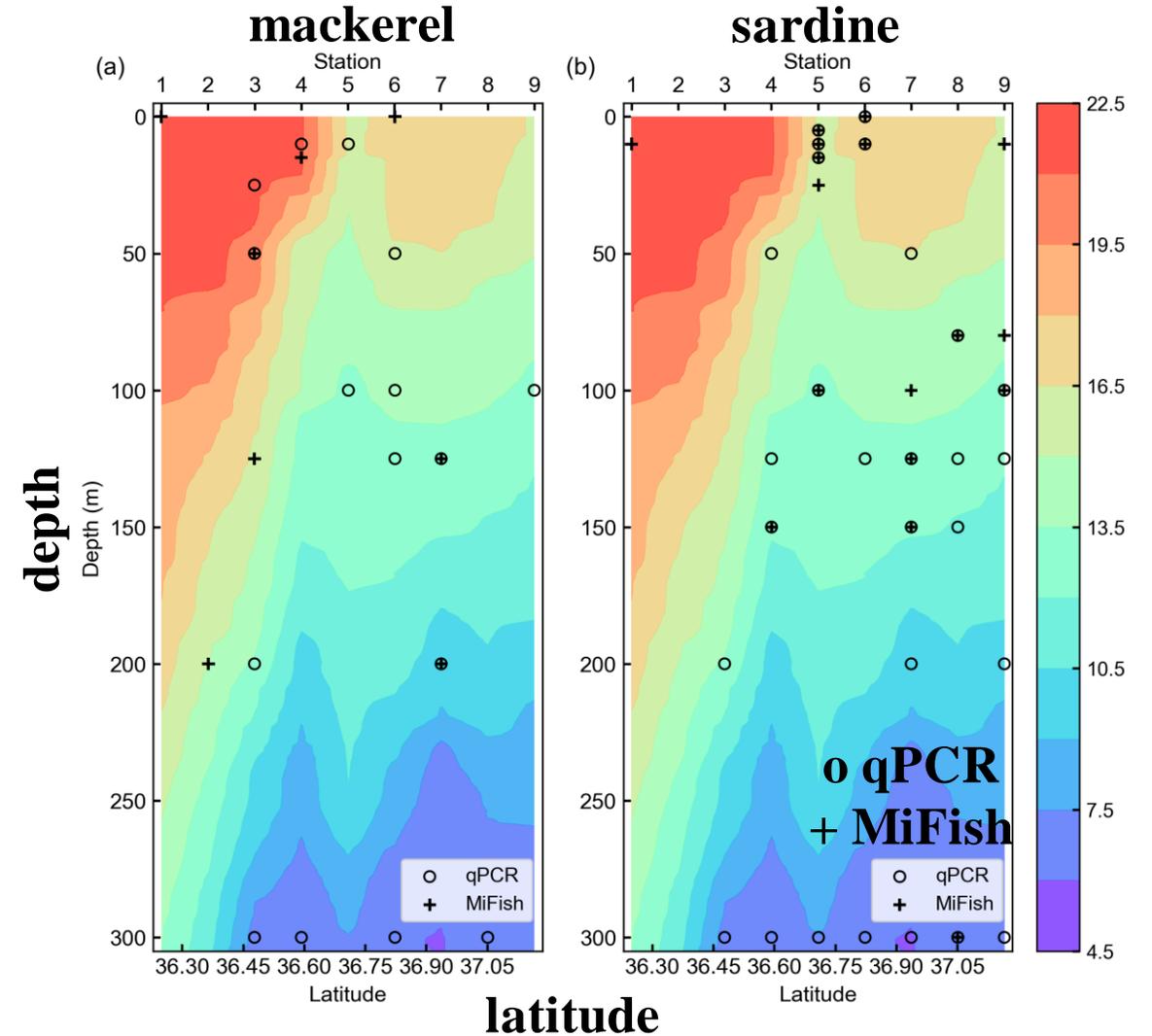
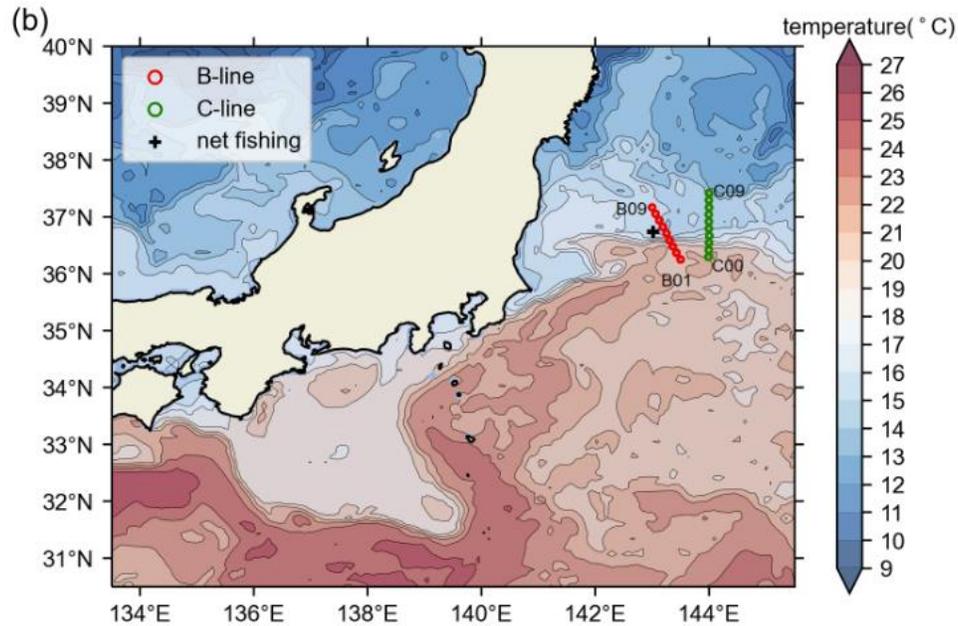
# OceanDNA



- Fish are releasing many types of creatures to the ocean water.
- Environmental DNA in the sea water (OceanDNA) can be used to detect fish distribution.
- Model derived fish migration accompanied with sub-mesoscale ocean structure can be observed used Ocean DNA.

Wong et al., Ito (2022, Environmental DNA)

# OceanDNA for distribution survey



High resolution fish distribution survey using OceanDNA is possible.

# Conclusion

- A new integrated method to elucidate environmental histories of larvae and juveniles, using natural tag chemical component analyses and fish growth–migration models, is proposed.
- The new integrated method can be used to elucidate the fish migration and life strategy and to build up improved fish growth-migration model.
- The improved model can be used to assess and project fish response to climate change.

**The new integrated approach will improve our understanding on climate impacts on marine ecosystem.**

**It will contribute to better sustainable use of marine ecosystem services.**

# References and contributors

- Guo C., S. Ito, Y. Kamimura, P. Xiu, accepted, Evaluating the influence of environmental factors on the early life history growth of chub mackerel (*Scomber japonicus*) using a growth and migration model. *Prog. Oceanogr.*
- Wong M. K., S. Nobata, S. Ito and S. Hyodo, 2022, Development of species-specific multiplex real time PCR assays for tracing the small pelagic fishes of North Pacific with environmental DNA. *Environmental DNA*, (online). <http://doi.org/10.1002/edn3.275>
- Harada Y., S. Ito, N. O. Ogawa, C. Yoshikawa, N. F. Ishikawa, M. Yoneda and N. Ohkouchi, 2022, Compound-specific nitrogen isotope analysis of amino acids in eye lenses as a new tool to reconstruct the geographic and trophic histories of fish. *Frontiers Mar. Sci.*, 8, Article 796532. <https://doi.org/10.3389/fmars.2021.796532>
- Guo C., S. Ito, M. Yoneda, H. Kitano, H. Kaneko, M. Enomoto, T. Aono, M. Nakamura, T. Kitagawa, N. C. Wegner, and E. Dorval, 2021, Fish specialize their metabolic performance to maximize bioenergetic efficiency in their local environment: conspecific comparison between two stocks of Pacific chub mackerel (*Scomber japonicus*). *Frontiers Mar. Sci.*, Article613965. <https://doi.org/10.3389/fmars.2021.613965>.
- Guo C., S. Ito, N. C. Wegner, L. N. Frank, E. Dorval, K. A. Dickson, D. H. Klinger, 2020, Metabolic measurements and parameter estimations for bioenergetics modelling of Pacific Chub Mackerel *Scomber japonicus*. *Fish. Oceanogr.*, 29, 215-226. <http://doi.org/10.1111/fog.12465>.
- Higuchi T., S. Ito, T. Ishimura, Y. Kamimura, K. Shirai, H. Shindo, K. Nishida, K. Komatsu, 2019, Otolith oxygen isotope analysis and temperature history in early life stages of the chub mackerel *Scomber japonicus* in the Kuroshio–Oyashio transition region. *Deep-Sea Res. II*, 169-170, Article 104660. <http://doi.org/10.1016/j.dsr2.2019.104660>.
- Ito S., T. Funamoto, O. Shida, Y. Kamimura, M. Takahashi, K. Shirai, T. Higuchi, K. Komatsu, T. Yokoi, T. Sakamoto, C. Guo, and T. Ishimur, 2018, A review of issues on elucidation of climate variability impacts on living marine resources and future perspective. *Oceanography in Japan*, 27, 59-73. [http://doi.org/10.5928/kaiyou.27.1\\_59](http://doi.org/10.5928/kaiyou.27.1_59).